

# DEVELOPING HPLC METHODS WHEN C18 COLUMNS DON'T WORK

Richard A. Henry, Technical Consultant, 983 Greenbriar Drive, State College, PA 16801 ([hyperLC@comcast.net](mailto:hyperLC@comcast.net))

Stephanie A. Schuster, William L. Johnson, and Conner W. McHale  
Advanced Materials Technology, Inc.  
3521 Silverside Road, Suite 1-K, Quillen Bldg.  
Wilmington, DE 19810

Thomas J. Waeghe, MAC-MOD Analytical, Inc.  
103 Commons Court  
Chadds Ford, PA 19317

Contact Dr. Stephanie Schuster for information ([SSchuster@advanced-materials-tech.com](mailto:SSchuster@advanced-materials-tech.com))

# CHOOSING PHASES OTHER THAN C18

- **Why choose a C18 phase?**
  - Very versatile and retentive for RPLC with simple binary gradients.
  - Proven stability with wide range of mobile phase and additives.
  - Low drift and bleed with LC-MS and other detectors.
  - Past success with C18 columns.
- **When to try another phase chemistry?**
  - Isomers or very similar analytes can't be resolved.
  - Too much retention or selectivity with C18 for desired analysis time.
  - Polar analytes not well retained with low or no organic modifier.
  - Polar analytes not well resolved even if retained.
  - A C18 method already in use is not rugged enough (revalidate).
- **Multiple column and solvent screening is recommended for resolving difficult mixtures and achieving rugged system performance.**

# FACTORS CONTROLLING HPLC/UHPLC RESOLUTION\*

$$R_s = \frac{\sqrt{N}}{4} \cdot \frac{k}{k+1} \cdot \frac{\alpha-1}{\alpha}$$

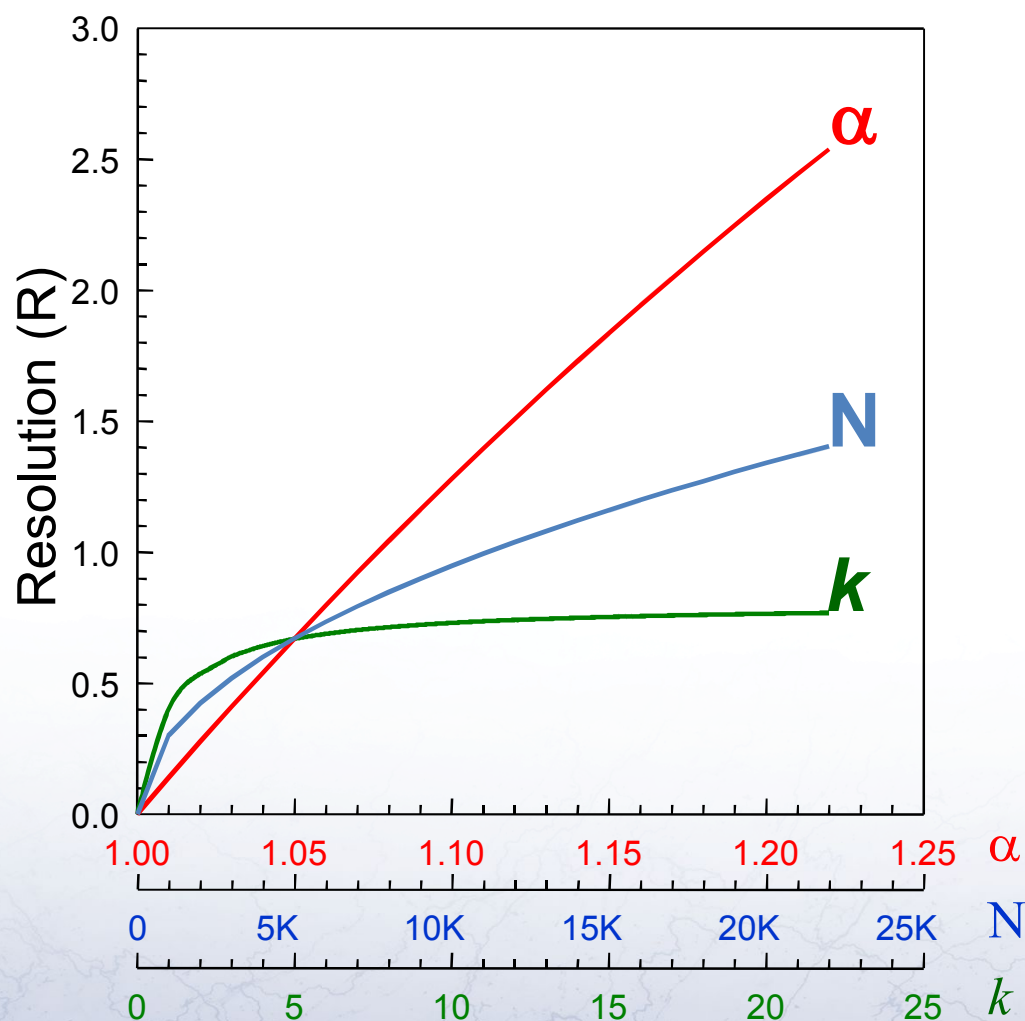
$$N = 16 (t_R/w_b)^2 \text{ or}$$

$$N = 5.5 (t_R/w_{1/2})^2$$

$$k = (t_R - t_0)/t_0$$

$$\alpha = k_2/k_1$$

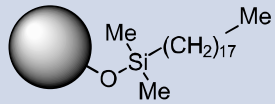
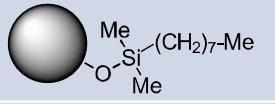
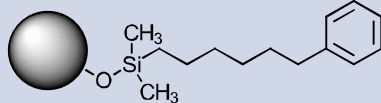
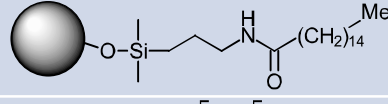
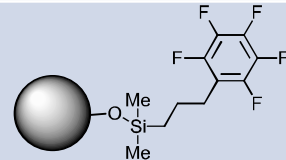
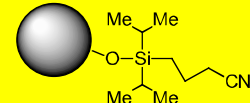
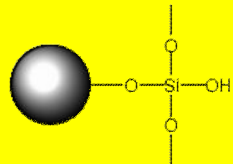
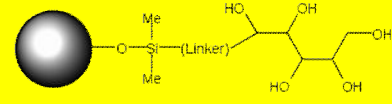
All factors are important, but selectivity is considered the most effective term to change.



\* Yun Mao, PhD Dissertation, Prof. Peter Carr, University of Minnesota, 2001.

# HALO<sup>®</sup> PHASES FOR SMALL MOLECULE SEPARATION

Industry  
Big Five

Phase	USP	Particle Sizes (μm)	Structure
<b>C18</b>	L1	2 2.7 5	
<b>C8</b>	L7	2 2.7 5	
<b>Phenyl-Hexyl</b>	L11	2 2.7 5	
<b>RP-Amide</b>	L60	2 2.7 5	
<b>PFP</b>	L43	2 2.7 5	
<b>ES-CN</b>	L10	2 2.7 5	
<b>HILIC (silica)</b>	L3	2 2.7 5	
<b>Penta-HILIC</b>	L95	2 2.7 5	

Phases For  
Special  
Situations

# FEATURES AND APPLICATIONS OF HALO PHASES

HALO Column	Features and Benefits	Best Applications
<b>C18, C8</b>	Excellent performance for broad range of analyte polarities	Analytes differing by an aliphatic or aromatic group
<b>Phenyl-Hexyl</b>	Complementary selectivity to alkyl phases Enhanced selectivity for aromatic compounds	Aromatic molecules with electron-withdrawing groups (NO <sub>2</sub> , COOH, COOR, halogens), heterocycles, benzodiazepines, highly aqueous conditions
<b>RP-Amide</b>	Complementary selectivity to alkyl phases Enhanced stability for minimum bleed and long life	Acidic and basic analytes, heterocycles, proton donors and acceptors, highly aqueous conditions
<b>PFP</b>	Complementary selectivity to alkyl phases Enhanced selectivity for stereoisomers Can be used in RPLC and HILIC modes	Basic analytes at low pH, stereoisomers, steroids, taxanes, substituted aromatics, highly aqueous conditions
<b>ES-CN</b>	Complementary selectivity to alkyl phases More retention for polar analytes and much less retention for non-polar analytes	Aromatic molecules with electron-withdrawing groups (NO <sub>2</sub> , COOH, COOR, halogens), heterocycles, benzodiazepines, highly aqueous conditions
<b>HILIC</b>	Can be used in HILIC and normal-phase modes	Enhanced sensitivity and peak shape for LC-MS, analyses of basic analytes in HILIC mode, normal-phase analysis with non-aqueous mobile phases
<b>Penta-HILIC</b>	Ideal for separation of highly polar compounds that are not retained in RPLC	Polar acids, bases and zwitterions that are not retained or are poorly retained with RPLC

# VARIABLES THAT AFFECT SELECTIVITY FOR RPLC METHOD DEVELOPMENT

## Column & Instrument System

- **Column (dictated by method)**
  - Stationary phase
  - Particle morphology
  - Particle composition
  - Particle pore size
- **Instrument (not dictated)**
  - Delay/Dwell volume

Methods will be easier to reproduce between labs if selectivity comes from the stationary phase with simple mobile phases. Operational variables contribute more to error because they are cumulative. Keep mobile phases as simple as possible.

## Operational (Mobile Phase)

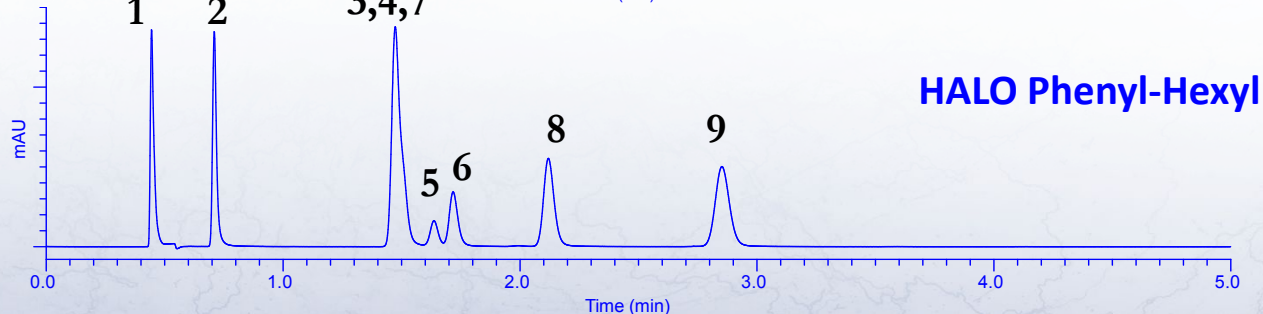
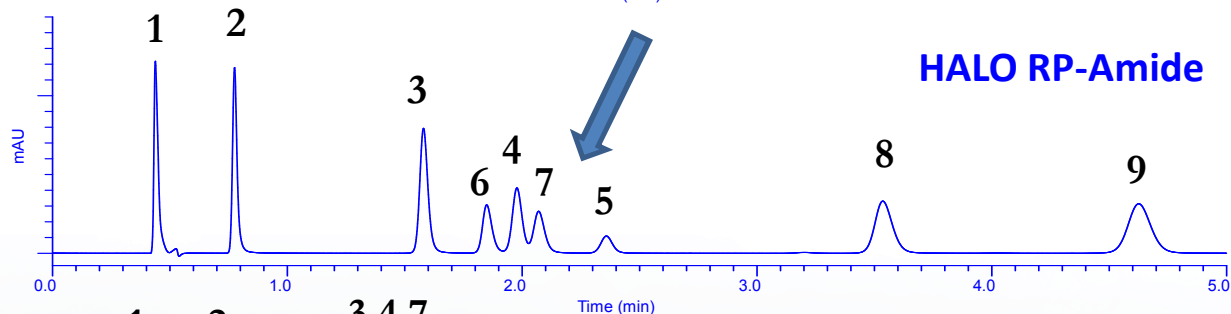
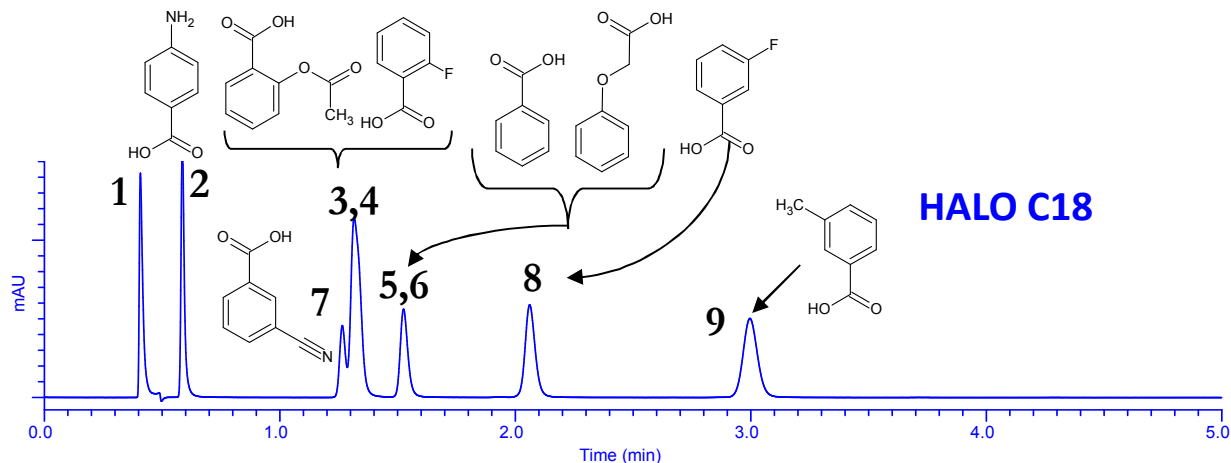
- Mobile phase pH
- Organic modifier choice
  - CH<sub>3</sub>CN, CH<sub>3</sub>OH, CH<sub>3</sub>CN/CH<sub>3</sub>OH blend, IPA, etc.
- % Organic (isocratic) or gradient slope (gradient)
- Column temperature
  - affects pH and ionization
- Additive type
  - TFA, H<sub>3</sub>PO<sub>4</sub>, phosphate, formate, acetate, NH<sub>4</sub>OH, etc.
- Additive concentration
  - ionic strength, ionization suppression

# RPLC SELECTIVITY RANKING VS HALO C18 AT DIFFERENT PH VALUES<sup>1-2</sup>

Phases Most Different from HALO C18	pH 2.8	pH 7
Most similar	C8	C8
	Phenyl-Hexyl	PFP
	ES-CN	Phenyl-Hexyl
	RP-Amide	ES-CN
Most different	PFP	RP-Amide

pH impacts charge on solutes that have acid-base properties and can also affect silica substrates. Silanol groups that are not covalently bonded will have neutral charge at low pH and negative charge at pH 7. Column rankings will also differ greatly between CH<sub>3</sub>CN and CH<sub>3</sub>OH and be very analyte dependent.

# PHASE COMPARISON FOR CARBOXYLIC ACID SELECTIVITY



HALO columns, 4.6 x 50 mm, 2.7  $\mu$ m

25:75 ACN/water with 0.1% formic acid, 25 °C, 1.0 mL/min

1. uracil
2. p-aminobenzoic acid
3. acetylsalicylic acid
4. 2-fluorobenzoic acid
5. benzoic acid
6. phenoxyacetic acid
7. 3-cyanobenzoic acid
8. 3-fluorobenzoic acid
9. m-toluic acid

H-bond acceptor (carbonyl) of **HALO RP-Amide** group attracts H-bond donor (acidic OH) and imparts more retention and better selectivity than either C18 or phenyl.

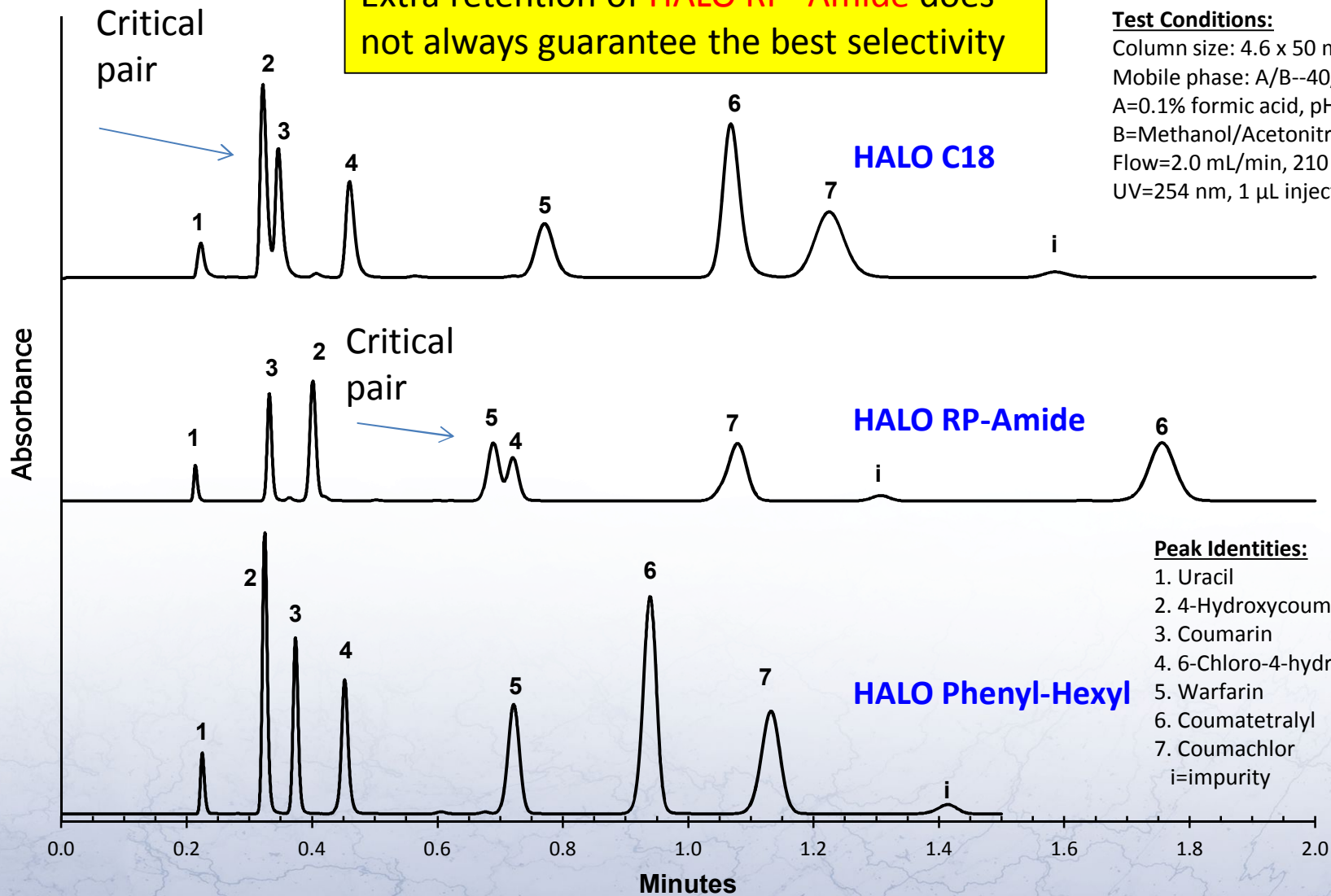


# FAST SEPARATION OF ANTICOAGULANTS ON HALO FUSED-CORE PACKINGS

Extra retention of HALO RP- Amide does not always guarantee the best selectivity

### Test Conditions:

Column size: 4.6 x 50 mm  
Mobile phase: A/B--40/60  
A=0.1% formic acid, pH=2.6  
B=Methanol/Acetonitrile-50/50  
Flow=2.0 mL/min, 210 Bar, 45 °C  
UV=254 nm, 1 µL injection



# PHASE COMPARISON FOR BENZODIAZEPINES

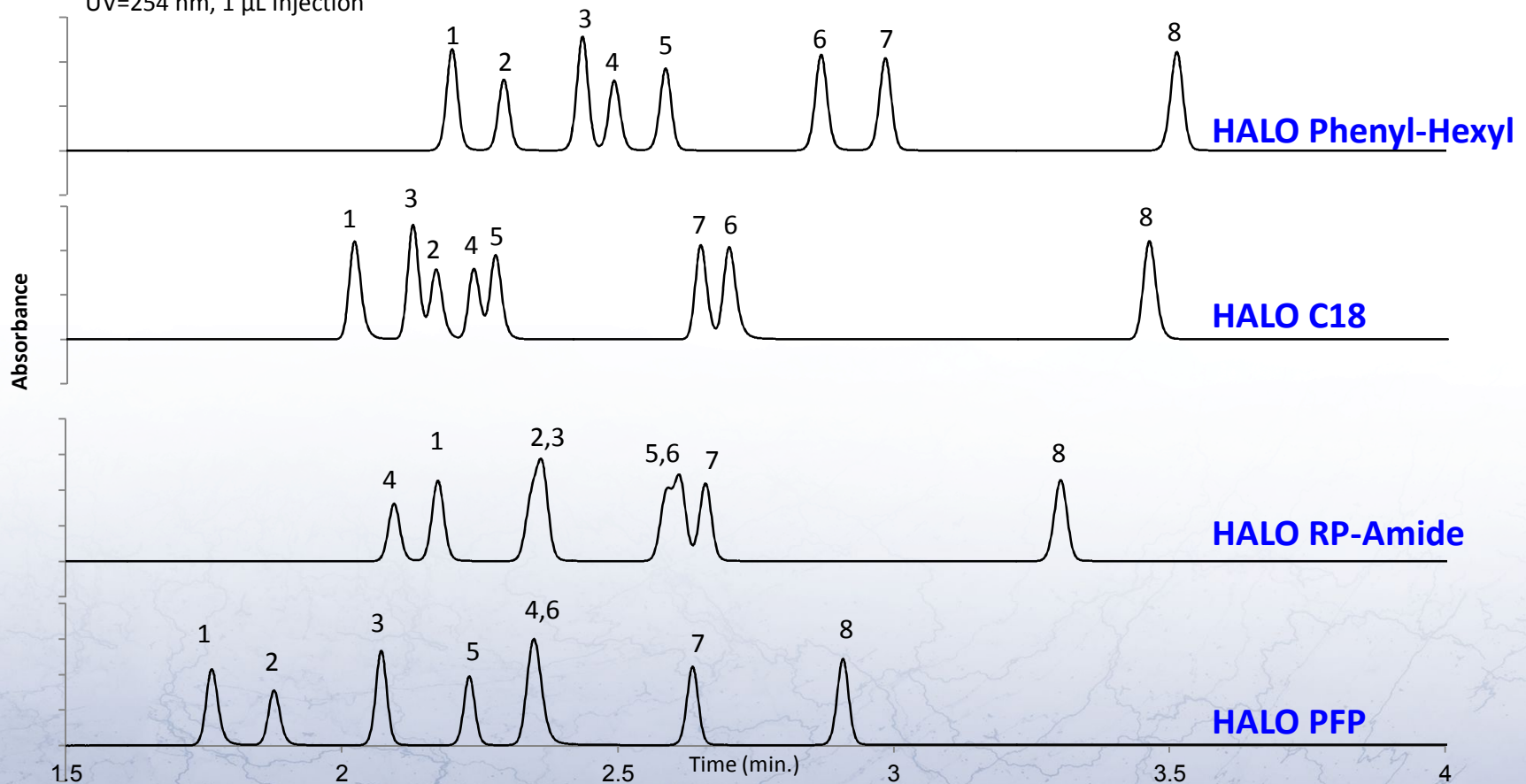
## Test Conditions:

Column size: 4.6 x 50 mm  
A = 25 mM Ammonium Acetate  
B = Acetonitrile  
Flow rate = 1.5 mL/min.  
Gradient = 34–63 %B in 3.5 min.  
Pressure = 200 bar  
Temperature = 35 °C  
UV=254 nm, 1 µL Injection

## Peak Identities:

- |               |                  |
|---------------|------------------|
| 1. Oxazepam   | 5. Clonazepam    |
| 2. Lorazepam  | 6. Temazepam     |
| 3. Nitrazepam | 7. Flunitrazepam |
| 4. Alprazolam | 8. Diazepam      |

**HALO Phenyl** phases are  $\pi$ -bases (electron-rich) and attract heterocyclic rings and other rings that are  $\pi$  acids (electron-poor) by shape-selective  $\pi$ - $\pi$  interaction.

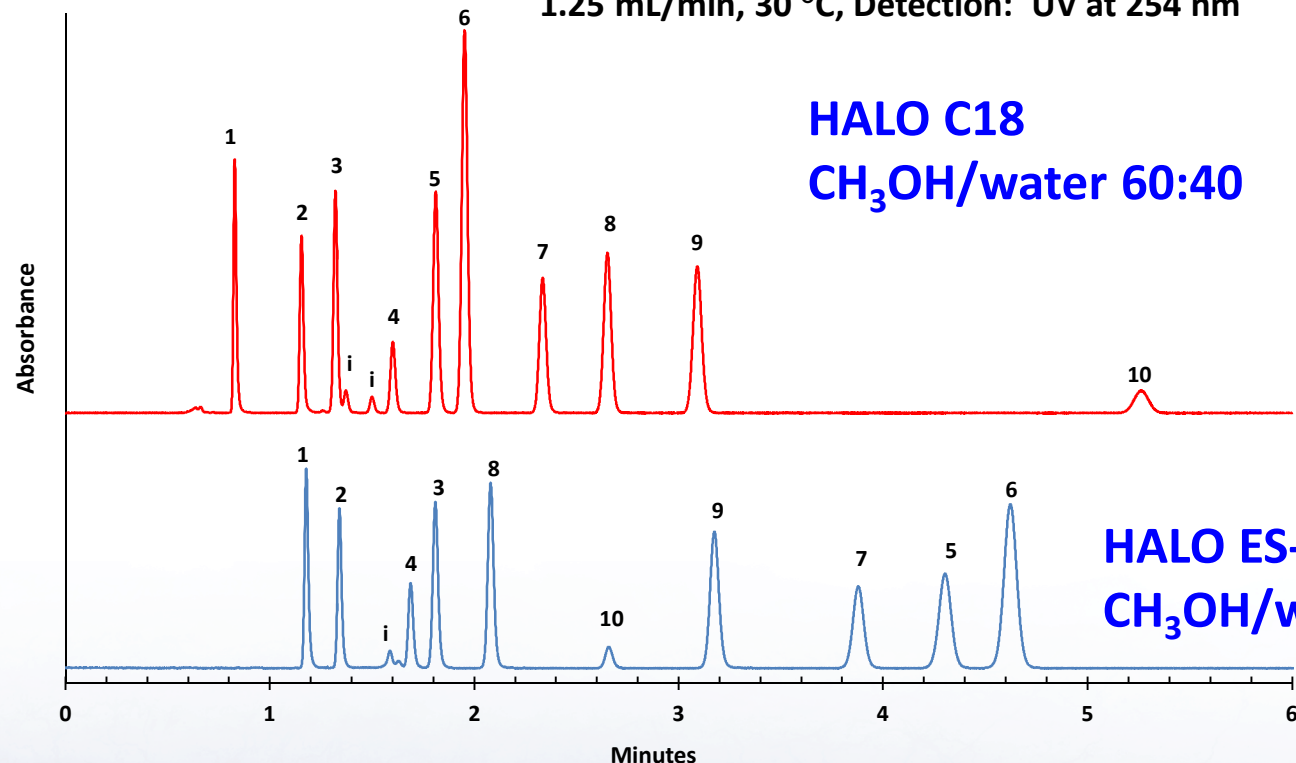


# CYANO PHASES OFFER ALTERNATIVE SELECTIVITY AND INCREASED RETENTION FOR POLAR ANALYTES

HALO columns, 4.6 x 100 mm, 2.7  $\mu\text{m}$   
1.25 mL/min, 30  $^{\circ}\text{C}$ , Detection: UV at 254 nm

## PEAK IDENTITIES:

1. Resorcinol
  2. Benzyl alcohol
  3. Phenylacetonitrile
  4. 1-Indanol
  5. 3,4-Dinitrotoluene
  6. 2,3-Dinitrotoluene
  7. 2,4-Dinitrotoluene
  8. Anisole
  9. 1-Chloro-4-nitrobenzene
  10. Toluene
- i = impurity



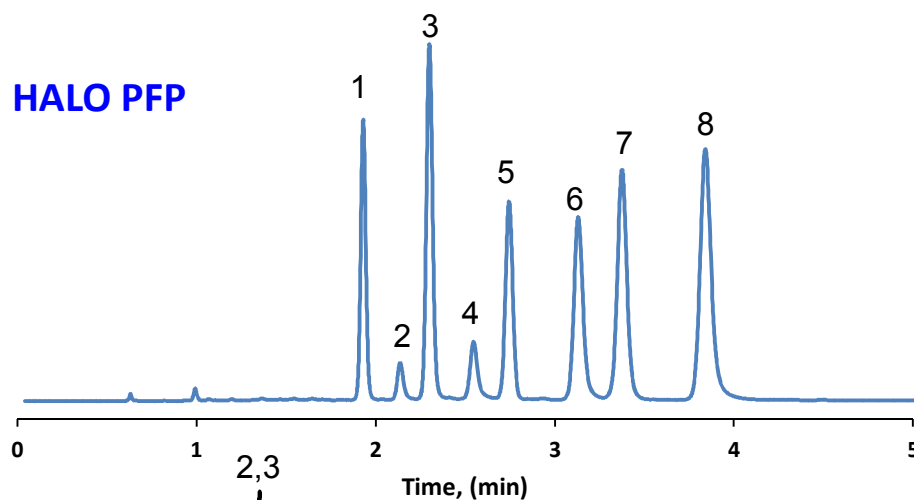
**HALO C18**  
**CH<sub>3</sub>OH/water 60:40**

**HALO ES-CN**  
**CH<sub>3</sub>OH/water 50:50**

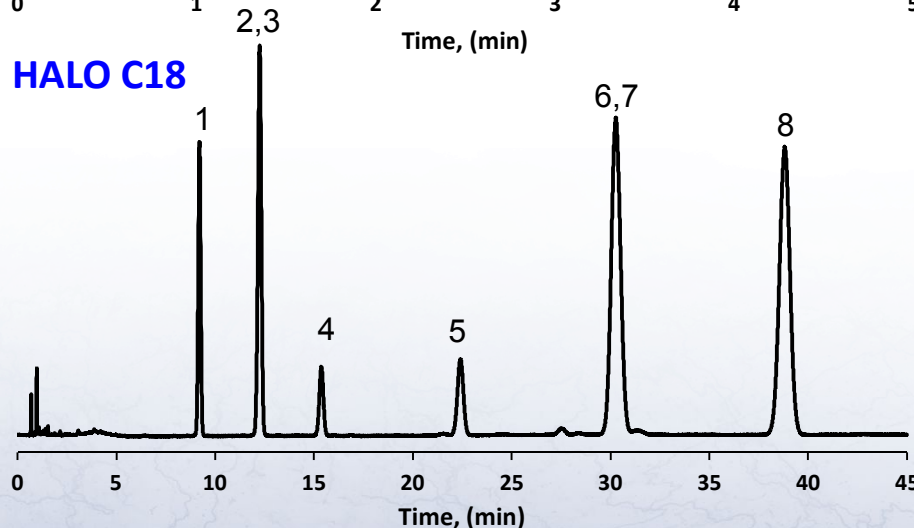
**HALO Cyano** phases interact by mild hydrophobic attraction plus dipole-dipole and dipole-aromatic forces and should be more popular in methods. Cyano requires only 10% less organic for equivalent retention, is very orthogonal to C18 for polar compounds, and is quite stable.

# FASTER SEPARATION AND BETTER RESOLUTION USING HALO PFP COLUMN VS. HALO C18

HALO PFP

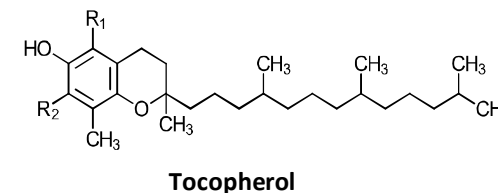
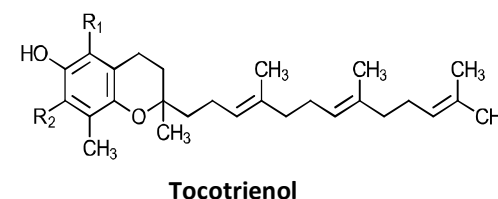


HALO C18



PEAK IDENTITIES:

1.  $\delta$ -tocotrienol
2.  $\beta$ -tocotrienol
3.  $\gamma$ -tocotrienol
4.  $\alpha$ -tocotrienol
5.  $\delta$ -tocopherol
6.  $\beta$ -tocopherol
7.  $\gamma$ -tocopherol
8.  $\alpha$ -tocopherol



HALO PFP phases are  $\pi$ -acids and can attract solutes by  $\pi$ - $\pi$  and several other mechanisms. HALO C18 has excess retention and selectivity overall, but cannot distinguish between isomer peaks 2,3 and 6,7.

HALO columns, 4.6 x 150 mm, 2.7  $\mu$ m

90:10 CH<sub>3</sub>OH/water, 25°C, 1.5 mL/min, Detection: Fluorescence: Excitation, 296 nm; Emission, 325 nm

# PHASE COMPARISON USING HILIC MODE

Nucleosides and nucleobases are often used as void markers for C18 columns, but they can be easily retained by Halo Penta-HILIC. Nucleosides, glycosides and other sugar-related compounds are ideal for **Halo HILIC** and **Penta-HILIC**. If RP and HILIC both show resolution, compare peak shape, equilibration, loading, and other factors.

## Test Conditions:

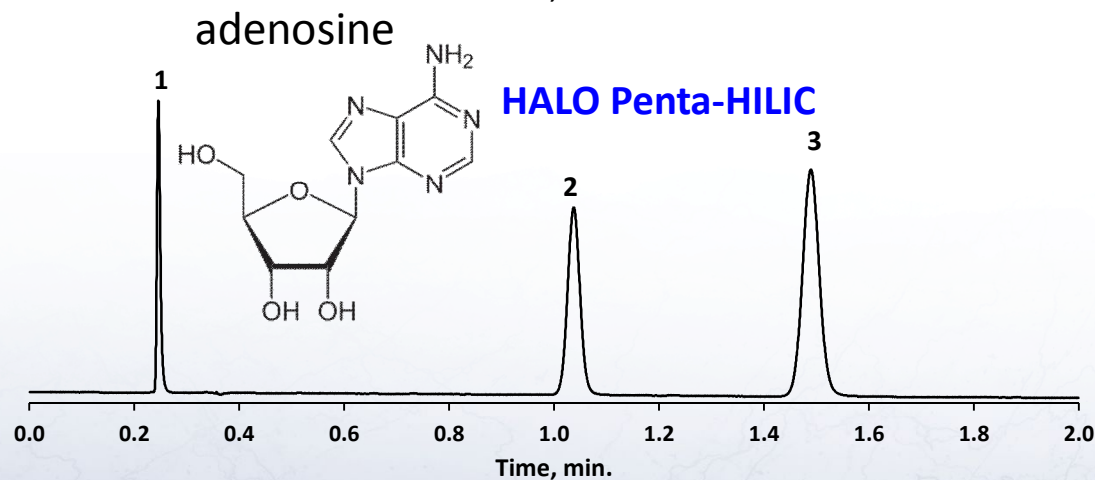
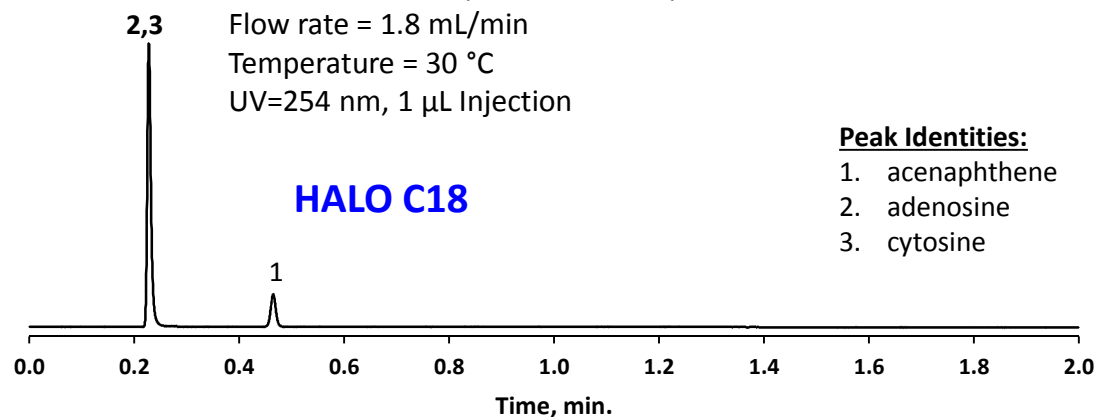
Column = 4.6 x 50 mm

Mobile Phase = 90/10 Acetonitrile/ 0.1 M ammonium formate buffer, pH = 3.0

Flow rate = 1.8 mL/min

Temperature = 30 °C

UV=254 nm, 1 µL Injection



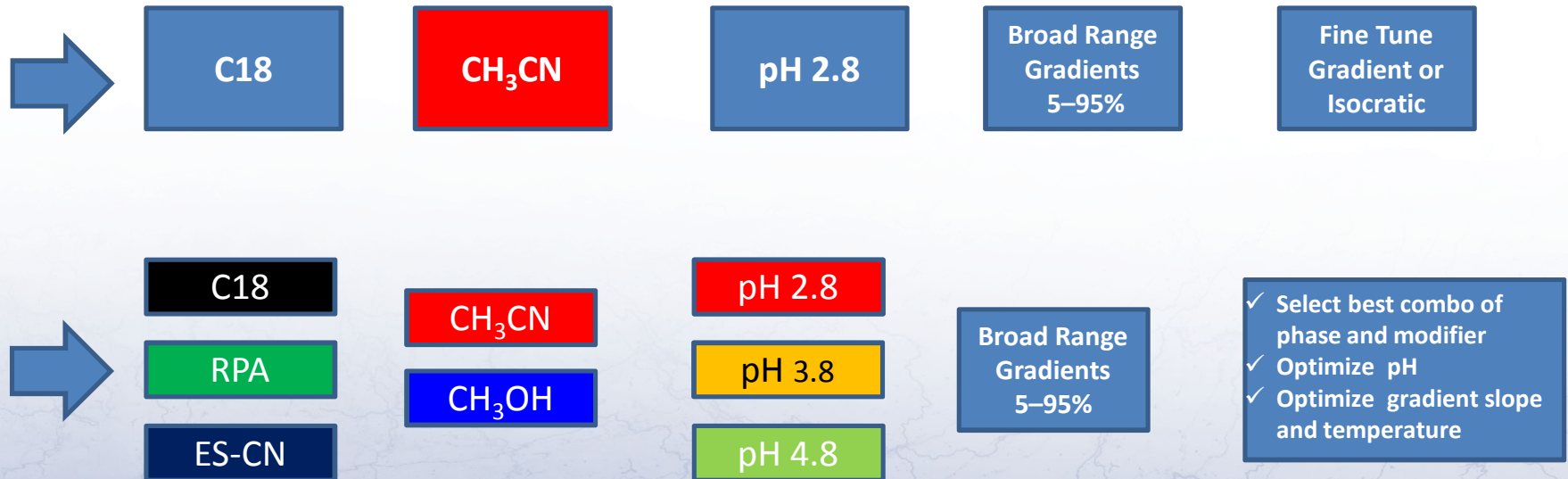
# DEVELOPING AN RPLC SEPARATION METHOD

Objective: to develop a good separation with the desired  $R_s$  and the necessary robustness for the desired purpose

- Method needs vary from relatively simple to very demanding
  - Simple: one lab for only several days or weeks
  - Moderate: one lab for several months
  - Demanding: method applied in multiple labs country-wide or world-wide and used for 5-10 years or more
- Sample complexity (known analytes) impacts method development time
  - < 5-6
  - < 10
  - > 10
- Robustness and ruggedness required for demanding methods
  - Robust- insensitive to small changes in variables
  - Rugged- reproducible during use by many labs, instruments, analysts, etc.

# METHOD DEVELOPMENT CAN VARY FROM RELATIVELY SIMPLE TO QUITE COMPLEX

- Separations that are simple often need little or no column screening so you can develop a separation quickly by varying solvent strength at low pH. Separations that are more difficult may not require a different phase, but you should compare different organic modifiers at multiple pH values.
- Demanding methods and more complex samples (e.g., stability-indicating method), usually require screening multiple phases, multiple organic modifiers, and several pH values to generate the most rugged separation.



# APPLICATION OF MULTIPLE PHASES FOR STABILITY INDICATING METHOD DEVELOPMENT

- Atorvastatin Calcium  
10 mg active/310 mg tablet
- Generate HCl-degraded and NaOH-degraded samples
- Pool acid- and base-treated samples together
- Compare five different HALO phases using both CH<sub>3</sub>CN and CH<sub>3</sub>OH at one pH (2.8, ammonium formate)
- Compare results and identify best option(s) for further development and optimization
- Use 3 x 50 mm, 2.7 μm HALO geometry
- Initially screened HALO C18 column using broad gradient with CH<sub>3</sub>CN (locate primary drug)
- Fine tuned to narrower ranges
- Compared all phases using narrower range using both CH<sub>3</sub>CN and CH<sub>3</sub>OH

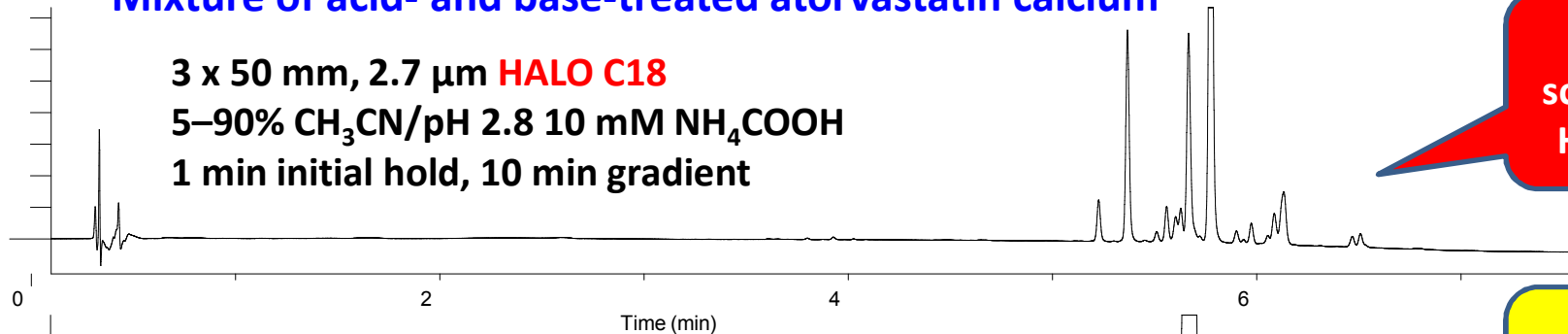


# BROAD RANGE GRADIENT WITH C18 IS USEFUL TO ESTABLISH HYDROPHOBICITY OF COMPLEX SAMPLES

## Mixture of acid- and base-treated atorvastatin calcium

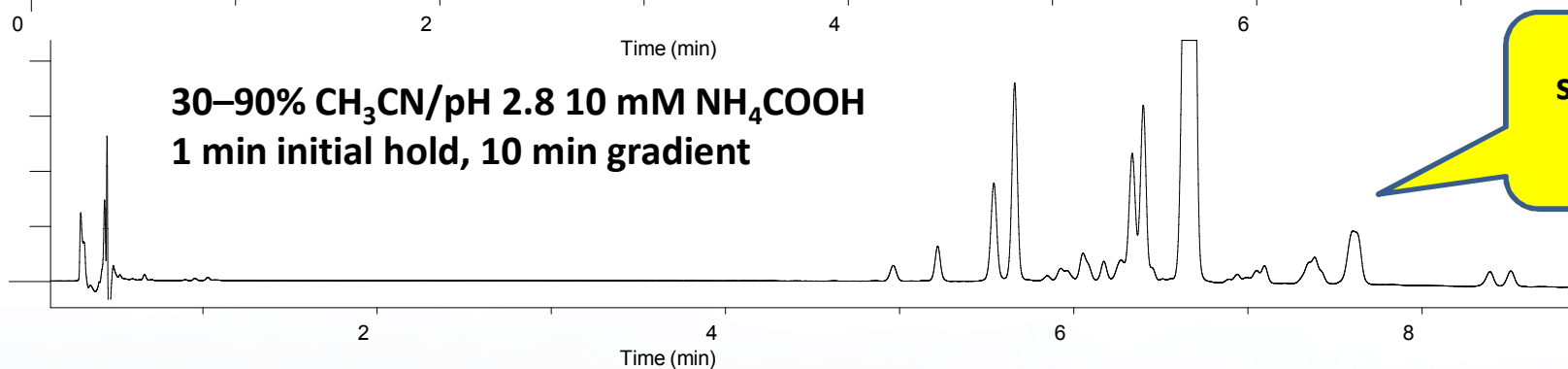
3 x 50 mm, 2.7  $\mu\text{m}$  HALO C18  
5–90%  $\text{CH}_3\text{CN}$ /pH 2.8 10 mM  $\text{NH}_4\text{COOH}$   
1 min initial hold, 10 min gradient

Initial  
screen with  
HALO C18



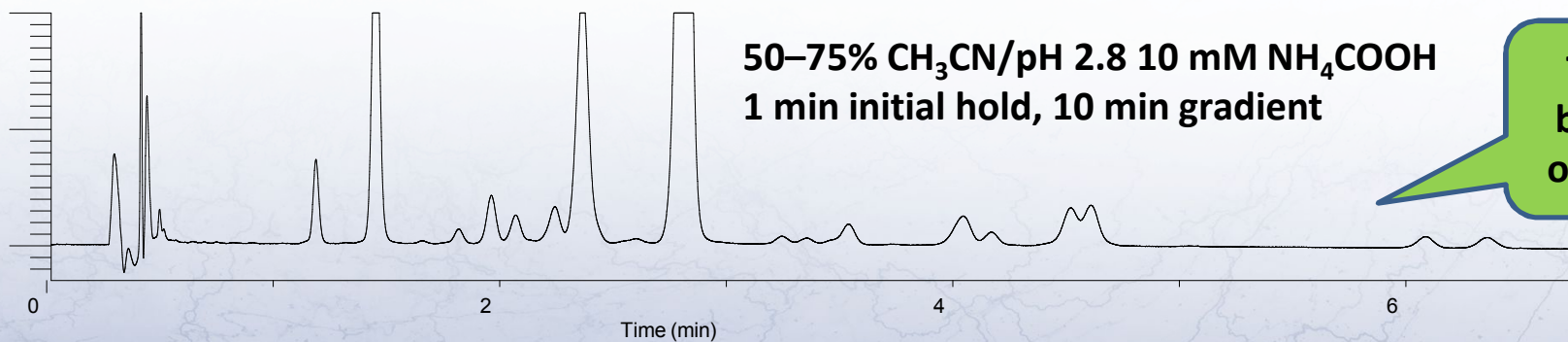
30–90%  $\text{CH}_3\text{CN}$ /pH 2.8 10 mM  $\text{NH}_4\text{COOH}$   
1 min initial hold, 10 min gradient

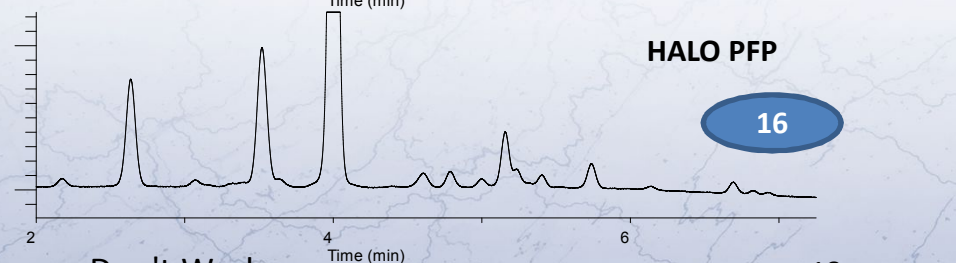
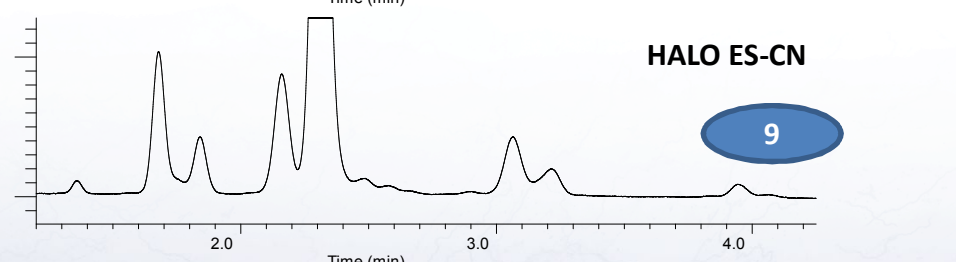
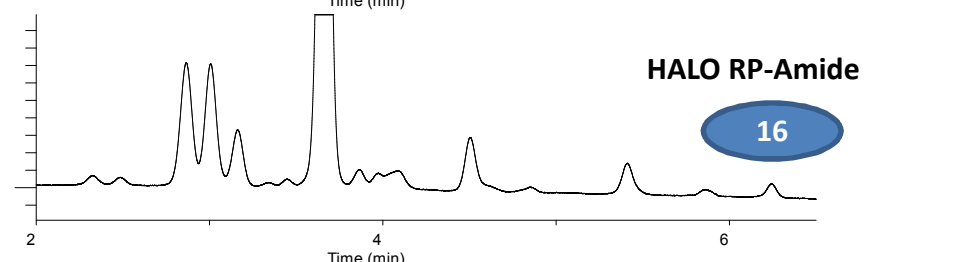
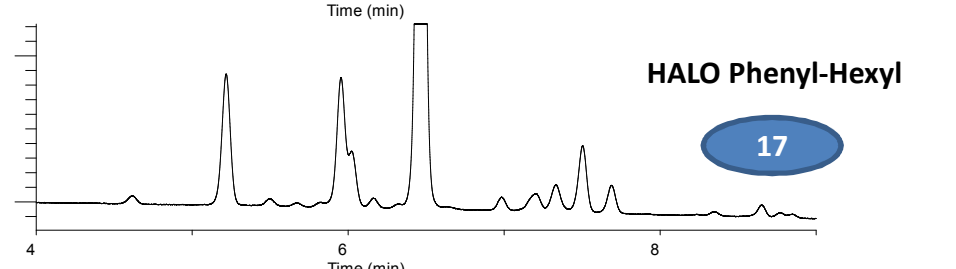
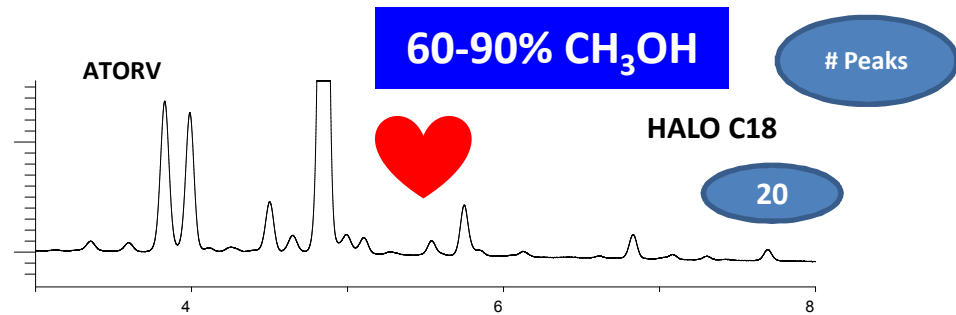
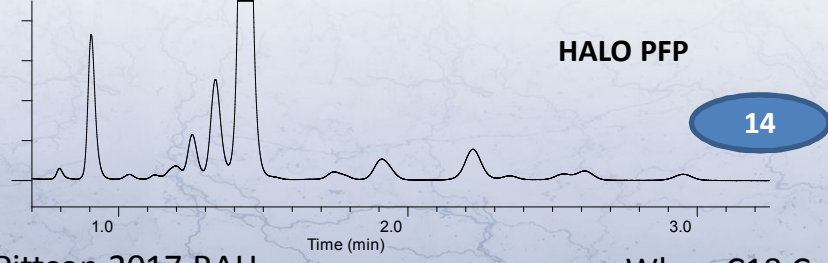
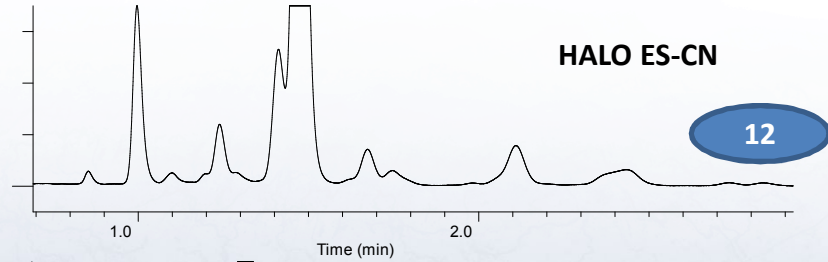
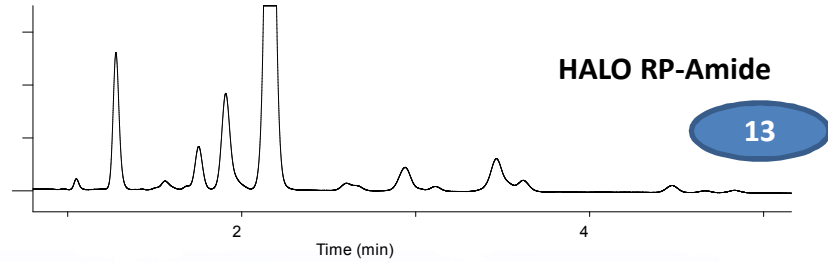
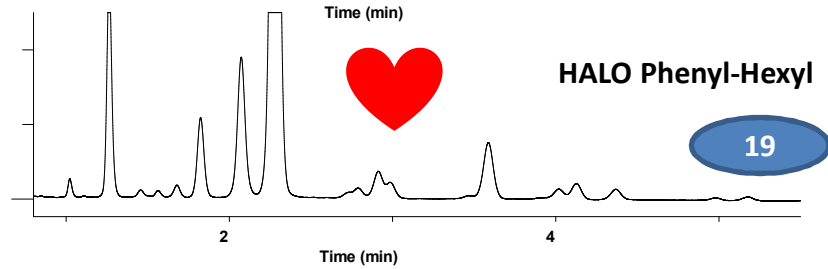
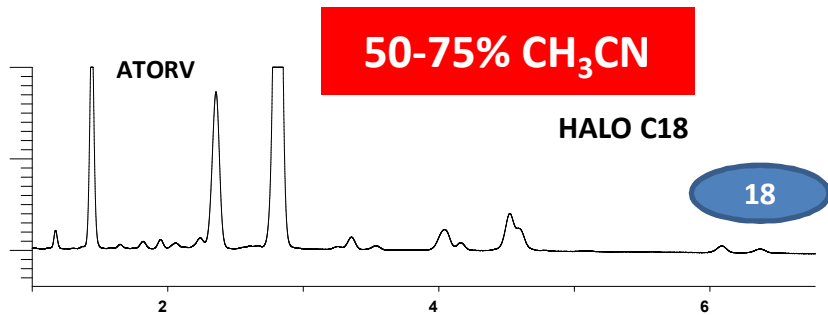
Raise  
starting %  
organic  
modifier



50–75%  $\text{CH}_3\text{CN}$ /pH 2.8 10 mM  $\text{NH}_4\text{COOH}$   
1 min initial hold, 10 min gradient

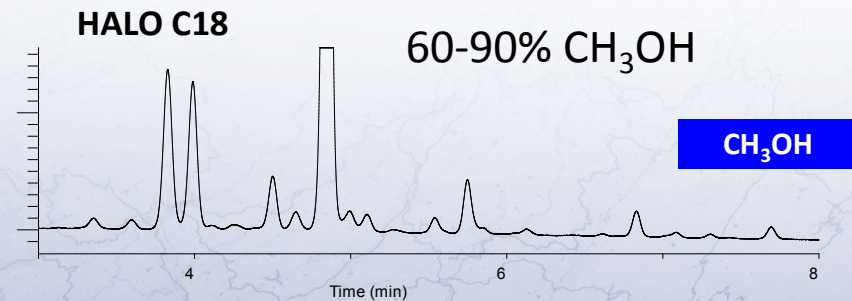
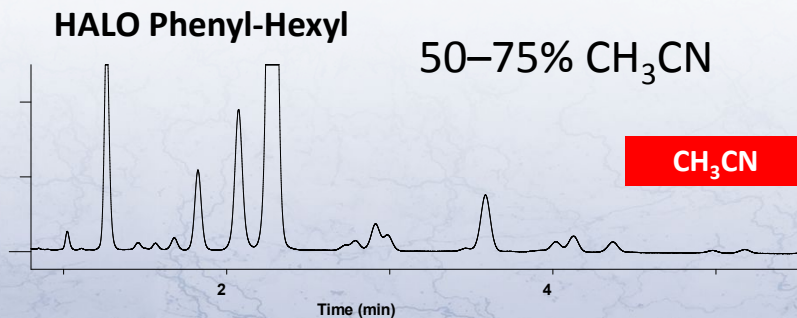
Truncate  
both ends  
of gradient





# HOW DO YOU CHOOSE WHICH COMBINATION TO DEVELOP AND OPTIMIZE FURTHER?

- Compare chromatograms for number of peaks observed.
- Compare shapes for detected peaks.
- Select most attractive phase/modifier combination(s) on points of merit:
  - # peaks separated
  - minimum  $R_s$  for critical peak pair
  - shortest analysis time
  - most peaks with acceptable USP  $T_f$
- If no clear winner emerges, run several gradients having differing slopes.
  - For example, 10 minutes and 25 minutes for Phenyl-Hexyl (50–75% ACN) and C18 (60-90%  $\text{CH}_3\text{OH}$ ).
  - Assess whether either combination stands out vs. criteria
- Compare separation and pressure on longer columns with higher efficiency.
- Check location of impurities for easier integration.



# SUMMARY AND RECOMMENDATIONS

- C18 columns are widely preferred in HPLC methods and may be satisfactory for most simple methods, but a C18 phase may not be the best solution for complex samples in demanding methods.
- Polar analytes can be separated better on phases that have greater polarity than C18, but selection of preferred phases and operating conditions to resolve complex mixtures may require column screening.
- A development strategy for demanding situations should be holistic and document all system parameters that control selectivity. This is the reason behind testing method variables for robust behavior before the method is validated. A robust method is likely to be rugged in use.
- The best approach for HPLC method development is to systematically screen multiple stationary phases and operating parameters. Special software and instrumentation (column, solvent selection) is available to assist with automated screening, selection, and optimization. Shortcuts can lead to fragile methods.

# REFERENCES AND ACKNOWLEDGEMENTS

1. Adapted from book, "Introduction to Modern Liquid Chromatography", 3rd Edition, L. R. Snyder, J. J. Kirkland and J. W. Dolan, 2010, p.29, Wiley & Sons.
2. "Orthogonal" separations for reversed-phase liquid chromatography; J. Pellett, P. Lukulay, Y. Mao, W. Bowen, R. Reed, M. Ma, R.C. Munger, J.W. Dolan, L. Wisley, K. Medwid, N.P. Toltl, C.C. Chan, M. Skibic, K. Biswas, K. A. Wells, and L. R. Snyder; Journal of Chromatography A, 1101 (2006) 122–135.
3. Column selectivity in reversed-phase liquid chromatography I. A general quantitative relationship; N.S. Wilson, M.D. Nelson, J.W. Dolan, L.R. Snyder, R.G. Wolcott and P.W. Carr; Journal of Chromatography A, 961 (2002) 171–193.
4. Column selectivity in reversed-phase liquid chromatography IV. Type-B alkyl-silica columns; J. J. Gilroy, J. W.
5. Dolan and L. R. Snyder; Journal of Chromatography A, 1000 (2003) 757–778.
6. <http://www.hplccolumns.org/>
7. <http://www.molnar-institute.com/drylab/> ("ColumnMatch").
8. D.V. McCalley and U.D. Neue, J. Chromatogr. A 1192, 225–229 (2008).
9. A.J. Alpert. Anal. Chem. 80, 62–76 (2008).
10. D.V. McCalley, J. Chromatogr. A 1171, 46–56 (2007).
11. A.J. Alpert et al., Anal. Chem. 82, 5253–5259 (2010).

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